Cholesteryl ester transfer protein and hyperalphalipoproteinemia in Caucasians

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Abstract It is unclear whether cholesteryl ester transfer protein (CETP) contributes to high density lipoprotein cholesterol (HDL-C) levels in hyperalphalipoproteinemia (HALP) in Caucasians. Moreover, even less is known about the effects of hereditary CETP deficiency in non-Japanese. We studied 95 unrelated Caucasian individuals with HALP. No correlations between CETP concentration or activity and HDL-C were identified. Screening for CETP gene defects led to the identification of heterozygosity for a novel splice site mutation in one individual. Twenty-five heterozygotes for this mutation showed reduced CETP concentration (-40%) and activity (-50%) and a 35% increase of HDL-C compared with family controls. The heterozygotes presented with an isolated high HDL-C, whereas the remaining subjects exhibited a typical high HDL-C/low-triglyceride phenotype. The increase of HDL-C in the CETP-deficient heterozygotes was primarily attributable to increased high density lipoprotein containing apolipoprotein A-I and A-II (LpAI:AII) levels, contrasting with an increase in both high density lipoprotein containing apolipoprotein A-I only and LpAI:AII in the other group. In This study suggests the absence of a relationship between CETP and HDL-C levels in Caucasians with HALP. The data furthermore indicate that genetic CETP deficiency is rare among Caucasians and that this disorder presents with a phenotype that is different from that of subjects with HALP who have no mutation in the CETP gene.—van der Steeg, W. A., G. K. Hovingh, A. H. E. M. Klerkx, B. A. Hutten, I. C. Nootenboom, J. H. M. Levels, A. van Tol, G. M. Dallinga-Thie, A. H. Zwinderman, J. J. P. Kastelein, and J. A. Kuivenhoven. Cholesteryl ester transfer protein and hyperalphalipoproteinemia in Caucasians. J. Lipid Res. 2007. 48: 674–682.

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In the general population, variations of high density lipoprotein cholesterol (HDL-C) levels are commonly

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ascribed to a combination of environmental and genetic factors (1). By contrast, extremely high levels of HDL-C [hyperalphalipoproteinemia (HALP)] are often the result of monogenetic disturbances. Among the genetic factors identified to date [such as scavenger receptor class B type 1 $(2, 3)$ and apolipoprotein C-III (apoC-III) deficiency (4)], deficiency of cholesteryl ester transfer protein (CETP) is a well-established cause of HALP (5). The CETP gene (6) is located on the long arm of chromosome 16 (7) and comprises 16 exons (8). In the circulation, CETP mediates the transfer of cholesteryl esters (CEs) from HDL to apoBcontaining lipoproteins (VLDL, intermediate density lipoprotein, LDL) (9, 10). Accordingly, human CETP deficiency is characterized by CE enrichment of HDL, which results in increased levels of HDL-C. This observation has led to the development of small molecular inhibitors of CETP that have been shown to increase plasma levels of HDL-C in humans (11–14).

The central role of CETP in human HDL metabolism only became evident after the identification of Japanese subjects with genetic CETP deficiency. Homozygotes for loss-of-function mutations showed up to four times increased HDL-C levels (15, 16). Since this discovery in 1989, CETP mutations have been identified primarily in Japan, and Maruyama et al. (17) showed that genetic CETP deficiency is in fact a very frequent cause of HALP in that country. Heterozygosity for a splice donor acceptor site mutation in intron 14 (Int14+1 G \rightarrow A) (15) and heterozygosity for a missense mutation in exon 15 (D442G) (16) were reported to underlie 74% and 62% increased

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Abbreviations: apoC-III, apolipoprotein C-III; BMI, body mass index; CE, cholesteryl ester; CETP, cholesteryl ester transfer protein; HALP, hyperalphalipoproteinemia; HDL-C, high density lipoprotein cholesterol; LDL-C, low density lipoprotein cholesterol; LpAI, high density lipoprotein containing apolipoprotein A-I only; LpAI:AII, high density lipoprotein containing apolipoprotein A-I and A-II.

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HDL-C levels compared with controls, respectively. Loss of CETP activity was further characterized by increased HDL2 cholesterol concentration (18) and increased mean HDL particle size (19). Given the central role of CETP in exchanging neutral lipids between HDL and apoB-containing lipoproteins, it is expected that CETP deficiency would also affect (V)LDL metabolism. However, heterozygotes for Intl4+1 G \rightarrow A or D442G have not been reported to exhibit changes in low density lipoprotein cholesterol (LDL-C), apoB, or triglyceride levels (5), despite an $\sim 50\%$ reduction of CETP activity levels. Regarding LDL particle size, it has been shown that the dimensions of these particles are distributed over an atypically wide range in subjects with homozygous CETP deficiency (20), but data covering this specific issue are scarce.

Together, genetic CETP deficiency and the use of CETP inhibitors have shown that CETP has a strong impact on HDL metabolism and that loss of CETP activity generates favorable changes of lipid profiles. Data regarding genetic CETP deficiency, however, are derived mainly from subjects living in Japan. In Caucasians, the literature on CETP and HALP is scarce. In fact, only a few Caucasian individuals with CETP deficiency have been described in case reports (21–24), and it is not known whether CETP plays a major role in determining HDL-C levels in non-Japanese subjects. Therefore, the first objective of this study was to investigate the relationship between CETP and HDL-C levels in Caucasian subjects with HALP. The second objective was to screen for genetic CETP deficiency in this cohort. The identification of a large family with a novel CETP gene defect allowed us to study in depth the impact of this mutation on lipid metabolism.

METHODS

Definition of the study group

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During the past decade, we have used our lipid clinic network and contacts with general practitioners in The Netherlands to collect plasma and DNA from individuals with HALP, with the intent to identify novel genes that control HDL-C levels. The vast majority of these individuals came to the attention of their physicians through general lipid tests that revealed high HDL-C levels. For this study, we examined 95 unrelated Caucasian index subjects (44 males, 51 females) from families in which HALP was established in at least three first-degree relatives. The average number of family members per index subject recruited was 14 and ranged from 4 to 342. Subjects were defined to have HALP if they presented with HDL-C levels above the 90th percentile for age and gender (for HDL-C percentile scale, see supplementary Table I) at two visits, without the presence of secondary causes that could lead to a HALP phenotype (extensive regular aerobic exercise, regular substantial alcohol intake, estrogen replacement therapy, or drugs such as fibrates, nicotinic acid, and phenytoin). Nine of the 95 subjects received statin therapy for suspected $(n = 1)$ or proven $(n = 4)$ cardiovascular disease or for unknown reasons $(n = 4)$. One of these subjects was additionally treated with a fibrate (gemfibrozil), but the presence of a HALP phenotype before the start of this therapy as well as the presence of this phenotype in three first-degree relatives were reasons for the inclusion of this individual.

Informed consent was obtained for blood sampling and genetic analyses, and the study was approved by the Medial Ethics Committee of the Academic Medical Center in Amsterdam, The Netherlands.

Genomic CETP DNA sequence analysis and mutation screening

To screen for genetic CETP deficiency, subjects with CETP concentrations $\leq 1.4 \mu g/ml$ (which is the lower limit of the normal range for CETP concentration of our ELISA) were selected from the HALP group. CETP gene sequence analysis was subsequently performed when low CETP concentrations in these subjects cosegregated with the HALP phenotype (defined above) in at least one family member. Genomic DNA was isolated from peripheral blood leukocytes. All 16 exons of the CETP gene, including intron-exon boundaries (minimum of 50 nucleotides into intronic DNA), and 1,500 bp of the CETP promoter were amplified by standard PCR using CETP-specific primers based on GenBank sequence NT_024766. Sequence reactions were performed using the Big Dye Terminator ABI Prism Kit on an Applied Biosystems model 310 automated DNA sequencer (Applied Biosystems, Nieuwerkerk a/d IJssel, The Netherlands).

Screening for the CETP-IVS7+1 mutation was performed by restriction fragment length polymorphism analysis using the forward 5'-CGGTGCCTGGTACACACTAG-3' and reverse 5'-CAT-AGTGCATCAGGTGGCTT-3' primers for PCR and digestion with XcmI (New England Biolabs, Beverly, MA), which digests wildtype DNA but not the mutant sequence.

RNA isolation and RT-PCR

Total RNA was isolated from peripheral leukocytes. One volume of whole blood was mixed with 9 volumes of lysis buffer $(155 \text{ mM NH}_4\text{Cl}, 10 \text{ mM KHCO}_3, \text{ and } 0.1 \text{ mM EDTA})$ and kept on ice for 15 min. Cells were collected by centrifugation at 3,000 g and washed twice by resuspension in lysis buffer and repeated centrifugation. The pellet was resuspended in Tripure, and RNA was isolated according to the manufacturer's instructions (Roche Diagnostics Nederland BV, Almere, The Netherlands). Firststrand cDNA was generated from 1 µg of total RNA with Superscript II Rnase H^- (Invitrogen, Breda, The Netherlands) with an oligo(dT) 14VN primer. Part of the cDNA reaction was used as template for a standard PCR. Primers used to detect alternative splicing products were 5'-TGGATCAAGCAGCTGTTCACA-3' and 5'-TGATGGGACTCCAGGTAGGA-3', sequences derived from exons 6 and 8, respectively. PCR products were visualized on ethidium bromide-containing agarose gels and, if needed, excised and prepared for sequencing.

To quantify alternatively spliced CETP mRNA, mRNA was isolated using Magnapure isolation (Roche Diagnostics Nederland BV) from abdominal adipose tissue obtained by needle aspiration from a heterozygote for CETP-IVS7 $+1$ and two control subjects. The amount of total CETP mRNA (wild type and mutant) and wild-type CETP mRNA was determined with Light-Cycler SYBR Green quantification using primer sets in exons 2 and 3 and exons 6 and 7, respectively. GAPDH mRNA levels were determined as an internal control.

Lipids, lipoproteins, apolipoproteins, and lipid-modifying proteins

All measurements were performed on fasting blood samples. Lipids were measured using standardized techniques. CETP concentration was determined by ELISA (25), and plasma apoA-I, apoA-II, apoB, and apoE were measured by nephelometry (Dade Behring, Marburg, Germany). High density lipoprotein containing apolipoprotein A-I only (LpAI) and high density lipoprotein containing apolipoprotein A-I and A-II (LpAI:AII) concentrations were determined by rocket gel electrophoresis (Sebia). Plasma CETP, LCAT, and phospholipid transfer protein activities were all measured using excess exogenous substrate methods (26). Activities are expressed as a percentage of the activity measured in pooled plasma obtained from 100 normolipidemic subjects. Lipoprotein subfraction concentrations and lipoprotein particle size were quantified by NMR spectrometry (27).

Statistical analysis

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Analyses were performed using the Statistical Program for the Social Sciences (version 12.0.2; SPSS, Inc., Chicago, IL). $P < 0.05$ was considered significant. Continuous variables with a skewed distribution were log-transformed before analysis.

Pearson correlation coefficients (r) were calculated to examine the relationship between CETP concentrations, CETP activity, specific CETP activity, and HDL-C. Linear regression analysis was performed to examine these relationships adjusted for age, sex, body mass index (BMI), smoking, and statin/fibrate use when these factors contributed significantly to the regression model. Results of the linear regression analyses are presented as standardized coefficients (β) .

A general linear model was used to compare lipids, (apo)lipoproteins, lipoprotein-modifying enzymes, and NMR data of the groups presented in Tables 1, 3, and 4. Using backward stepwise linear regression analysis, we again adjusted for age, sex, BMI, smoking, and statin use when these parameters contributed significantly to the model. To investigate the effect of fibrate therapy, analyses were also performed after exclusion of the individual receiving gemfibrozil.

RESULTS

CETP and HALP

The first objective of this study was to investigate the relationship between CETP and HDL-C levels in Caucasian subjects with HALP. To this purpose, we studied 95 unrelated probands from families in which HALP was established in at least three first-degree relatives. The average HDL-C level in this group was 2.35 ± 0.42 mmol/l (Table 1), with higher HDL-C levels in women compared with men (2.61 \pm 0.32 vs. 2.06 \pm 0.33 mmol/l; P < 0.0001) (Table 1). Plasma CETP concentration was normally distributed and averaged 1.89 ± 0.60 µg/ml. Males presented with nonsignificantly lower CETP concentrations compared with females (1.79 \pm 0.54 vs. 1.98 ± 0.60 μ g/ml; $P = 0.21$) (Table 1). Mean plasma CETP activity in the entire group was similar to pooled plasma of healthy volunteers (99.0% vs. 100%). Mean CETP activity was slightly lower in men $(96.8 \pm 19.4\%)$ compared with women (100.9 \pm 29.7%), but this difference was not statistically significant ($P = 0.42$). Thus, compared with men, the women presented with higher HDL-C levels accompanied by nonsignificant increases in CETP concentration and activity (exclusion of the individual receiving fibrate therapy generated similar results).

Using the data for the entire group, we identified a correlation between CETP concentration and CETP activity $(r = 0.67, P < 0.0001)$ (Table 2). Addressing the first question of this study, HDL-C levels were not correlated with CETP activity $(r = 0.17, P = 0.16)$, CETP concentration ($r = 0.14$, $P = 0.17$), or specific activities of CETP ($r = -0.17$, $P = 0.16$). Linear regression analysis, adjusted for age, sex, BMI, smoking, and statin/fibrate use when necessary, generated similar results. CETP activity was assessed in only 71 of 95 subjects, as a result of lack of sufficient plasma.

Identification of a novel splice site mutation in the CETP gene in one family

Our second objective was to screen for genetic CETP deficiency in our cohort. To this purpose, we selected 13 subjects with CETP concentrations $\leq 1.4 \mu g/ml$ (which is the lower limit of the normal range for CETP concentration of our ELISA). CETP gene sequence analysis was performed when low CETP concentrations in these subjects cosegregated with the HALP phenotype in at least one family member [this was the case in 12 (8 men, 4 women) of the 13 individuals with low CETP concentrations]. This analysis revealed a number of CETP gene variations (listed in supplementary Table II) that were either previously described polymorphisms (28, 29) or were not expected to affect CETP gene transcription, based on their positions in

TABLE 1. Demographic and clinical characteristics, lipids, CETP concentration, CETP activity, and specific CETP activity in 95 individuals with HALP

All $(n = 95)$	Men $(n = 44)$	Women $(n = 51)$	\boldsymbol{P}
57 ± 14	57 ± 14	58 ± 14	
20.0	20.5	19.6	
24.0 ± 2.7	24.2 ± 2.6	23.8 ± 2.8	
6.31 ± 1.01	6.25 ± 1.05	6.37 ± 0.98	
3.58 ± 0.98	3.80 ± 0.97	3.39 ± 0.97	
2.35 ± 0.42	2.06 ± 0.33	2.61 ± 0.32	< 0.0001
0.83 ± 0.39	0.85 ± 0.49	0.81 ± 0.29	
1.89 ± 0.60	1.79 ± 0.54	1.98 ± 0.60	0.21
99.0 ± 25.4	96.8 ± 19.4	100.9 ± 29.7	0.42
54.3 ± 12.8	56.9 ± 13.2	52.0 ± 12.2	0.11

BMI, body mass index; CETP, cholesteryl ester transfer protein; HALP, hyperalphalipoproteinemia; HDL-C, high density lipoprotein cholesterol; LDL-C, low density lipoprotein cholesterol. Values are given as means \pm SD; P values are for men versus women after correction for differences in age, sex, smoking, BMI, and statin use when these parameters contributed significantly to the model. Exclusion of the individual receiving fibrate therapy did

not affect the outcome.
^a Assessed in 71 of 95 subjects (males, 33 of 44; females, 38 of 51) without selection bias.

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http://www.jlr.org/content/suppl/2007/01/17/M600405-JLR20
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TABLE 2. Relationships between CETP activity, CETP concentration, specific CETP activity, and HDL-C in 95 subjects with HDL-C levels above the 90th percentile for age and sex

Variable		P1		P ₂
CETP activity ^{a} and CETP concentration		$0.67 \le 0.0001$		$0.67 \le 0.0001$
CETP activity ^{a} and HDL-C	0.17	0.16	0.05	0.66
CETP concentration and HDL-C	0.14	0.17	0.11	0.30
Specific CETP activity ^{<i>a</i>} and HDL-C -0.17		0.16	-0.17	0.16

 r , Pearson correlation coefficient; P1, level of significance; β , standardized coefficient derived from linear regression analysis adjusting for age, sex, smoking, BMI, and statin/fibrate use when these parameters contributed significantly to the model; P2, level of significance.
^{*a*} Assessed in 71 of 95 subjects (males, 33 of 44; females, 38 of 51)

without selection bias.

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nonregulatory promoters or intronic sequences. We did not analyze these variations further, but focused on one woman who turned out to be heterozygous for a novel splice site mutation. It concerned a $G \rightarrow T$ nucleotide substitution at nucleotide $+1$ of intron 7 (CETP-IVS7+1). This sequence variation was not present in the remaining 94 probands with high HDL-C or in 200 normolipidemic unrelated control subjects.

We recruited 190 family members of this CETP-IVS7+1 proband and screened for this defect by PCR (see Methods for details). This led to the identification of 24 additional heterozygotes, but no homozygotes were found. Supplementary Fig. I shows the most relevant part of the pedigree of this family, and gender, age, mutation carrier status, HDL-C levels, and HDL-C percentiles are listed in supplementary Table III. The penetrance of the CETP-IVS $7+1$ mutation with respect to CETP concentration and HDL-C level was very high: 23 of 25 carriers (92%) had CETP concentrations $\langle 1.4 \mu g/ml \rangle$ (range, 0.58–1.18 $\mu g/ml$), and 16 of 25 subjects (68%) had HDL-C above the 90th percentile (for percentile scale, see supplementary Table I).

Molecular pathology of the CETP-IVS $7+1$ mutation

The CETP-IVS7 $+1$ mutation disrupts the consensus splice donor site sequence of intron 7 (GT \rightarrow TT). To delineate the molecular consequences of this mutation, we analyzed CETP mRNA isolated from peripheral white blood cells of an affected individual and a control subject. Using RT-PCR, control mRNA produced a single 212 bp fragment, indicating proper mRNA processing (Fig. 1A). However, an additional aberrant PCR product of \sim 150 bp was identified when using mRNA of the CETP-IVS7 $+1$ carrier. DNA sequence analysis of this PCR product showed that the sequence of exon 6 was followed directly by exon 8, indicating that the mutation caused exon 7 to be skipped (61 bp) (Fig. 1B). Because of the resulting frameshift, translation of the aberrant mRNA is predicted to result in a truncated CETP protein of 196 amino acids (Fig. 1C). Using quantitative PCR, we assessed the concentration of the two CETP mRNA variants in adipose tissue of the affected individual. The estimated amount of mutant mRNA was 24.7 (arbitrary units), compared with a total of 93.3 (arbitrary units) of wild-type mRNA (both compared with GAPDH expression levels). These results

Fig. 1. Molecular characterization of the cholesteryl ester transfer protein (CETP)-IVS7+1 defect. A: RT-PCR of RNA from a CETP-IVS711 carrier and a control subject. Plasmid CETP cDNA was used as a control. Primers were located in exon 6 and exon 8. Control mRNA produces a 212 bp fragment indicating proper processing; an additional fragment is present in mRNA of the IVS711 carrier. B: Partial DNA sequence of the mutant RT-PCR product $(151 bp)$ of the IVS7+1 carrier showing the skipping of exon 7. C: Hypothetical effects of the frameshift caused by exon skipping on translation of the mutant CETP mRNA. Values in parentheses indicate total lengths of wild-type (wt) and mutant (mut) CETP proteins in amino acids.

indicate that 20% of the pool of CETP mRNAs consisted of the alternatively spliced mRNA product.

The ELISA used to determine plasma CETP concentration did not allow for the detection of the hypothesized truncated CETP protein, because it lacks the epitopes for the TP1 and TP2 antibodies that were used. To address whether this truncated product was synthesized and present in plasma of CETP-IVS7+1 carriers, another ELISA using monoclonal antibody ScFv 1CL8 directed against amino acids 68–87 and 110–129 of CETP was used (30). Data obtained with this antibody were not different Supplemental Material can be found at:
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from those obtained in the first assay, suggesting that there is no detectable truncated CETP protein present in fasted plasma of the carriers of this mutation. These data thus suggest that the CETP-IVS7+1 heterozygotes only have wild-type CETP protein in their circulation.

Heterozygosity for CETP-IVS7+1: effects on CETP and HDL-C

To carefully study the effects of the mutation in this family, the 25 heterozygotes for the CETP-IVS7+1 mutation were matched to 25 family controls of similar age, sex, and BMI (selected from 190 family members that were recruited). These 25 family controls presented with a lipid profile that was similar to that of a cohort of 2,381 healthy normolipidemic subjects from our database (for more information, see supplementary Table IV). This finding provided the basis for the use of the 25 family members as a control group in the analysis described below. The rationale for using these family controls is that lifestyle and genetic background affecting lipid parameters can be assumed to be similar in the family controls compared with those in family members carrying the CETP mutation. Three heterozygotes used statin therapy for proven cardiovascular disease; none of the control subjects received lipid-lowering medication. Illustrating the penetrance of this mutation, the heterozygotes presented with an average 40% decrease of plasma CETP concentration (0.94 ± 0.28) vs. 1.58 ± 0.31 μ g/ml; $P < 0.0001$) (**Table 3**) and a concomitant mean 50% reduction in plasma CETP activity $(40.1 \pm 12.4 \text{ vs. } 81.1 \pm 20.4\%; P \leq 0.0001)$ (Table 3) compared with family controls. Affected males $(n = 13)$ and females $(n = 12)$ showed nearly identical reductions of CETP activity and CETP concentration levels $(40.0 \pm 6.6 \text{ vs.})$ $40.3 \pm 18.9\%$ and 0.96 ± 0.30 vs. 0.93 ± 0.26 μ g/ml, respectively). Compared with family controls, HDL-C levels were strongly increased in the heterozygotes (2.06 ± 0.65 vs. $1.52 \pm 0.39 \, \text{mmol/l}; P \leq 0.0001$ (Table 3). Thus, in

contrast with our finding in the entire HALP cohort that CETP was not associated with HDL-C levels, these data show that a profound loss of CETP activity as a result of partial CETP deficiency can underlie HALP.

Lipid metabolism in CETP-IVS7 $+1$ heterozygotes, family controls, and 94 individuals with HALP

Next, we compared the 25 heterozygotes for CETP- $IVS7+1$ with individuals with HALP in which no CETP deficiency $(n = 94)$ was identified. Although we sequenced the CETP gene only in individuals with low CETP concentrations, we here assumed the absence of CETP gene defects in the remaining subjects with CETP concentrations and CETP activity levels in the normal range. We used backward stepwise linear regression analysis, in which we adjusted for age, sex, BMI, smoking, and statin use when necessary. Exclusion of the individual receiving fibrate therapy did not affect the outcome. In the group with HALP $(n = 94)$, HDL-C levels were significantly higher compared with the CETP-IVS7+1 heterozygotes (2.35 \pm 0.43 vs. 2.06 \pm 0.65 mmol/l; $P = 0.04$). Compared with controls, the increase of HDL-C levels in the HALP group was reflected by increases of both LpAI and LpAI:AII levels. However, the increase in LpAI was larger $(+0.24 \text{ g/l})$ compared with that of LpAI:AII $(+0.13 \text{ g/l})$. By contrast, heterozygosity for the $CETP-IVS7+1$ mutation had little and no significant effect on LpAI levels $(0.57 \pm 0.20 \text{ vs. } 0.51 \pm 0.13 \text{ g/l} \text{ in controls};$ $P = 0.23$) but had a more profound and statistically significant effect on LpAI:AII levels (1.30 \pm 0.25 vs. 1.11 \pm 0.20 g/l in controls; $P = 0.003$). Although apoE concentration appeared higher in the HALP and CETP-IVS+1 groups, this did not reach statistical significance.

The HALP group further exhibited 40% lower triglyceride levels compared with the CETP-IVS7+1 heterozygotes (0.83 \pm 0.39 vs. 1.37 \pm 0.98 mmol/l; P < 0.0001) and higher LDL-C levels $(3.58 \pm 0.99 \text{ vs. } 2.80 \pm 1.02 \text{ mmol/l})$; $P = 0.03$). By contrast, apoB levels were not different be-

TABLE 3. Demographic and clinical characteristics, plasma CETP concentration and activity, lipids, and apolipoproteins in unrelated subjects with HALP, heterozygotes for a novel CETP splice site mutation $(IVS7+1)$, and family controls

Variable	Family Controls $(n = 25)$	CETP-IVS7+1 Heterozygotes $(n = 25)$	HALP $(n = 94)$	P1	P ₂	P ₃
Sex $(\%$ male)	54.2	52	46.8			
Age (years)	40 ± 22	39 ± 22	57.4 ± 13.8			
Smoking $(\%)$	19.0	20.0	19.1			
BMI (kg/m^2)	24.3 ± 5.1	23.4 ± 4.9	24.0 ± 2.7			
CETP concentration $(\mu g/ml)$	1.58 ± 0.31	0.94 ± 0.28	1.90 ± 0.58	< 0.0001	0.01	< 0.0001
CETP activity $(\%)$	81.1 ± 20.4	40.1 ± 12.4	99.9 ± 24.3	< 0.0001	0.005	< 0.0001
$HDL-C$ (mmol/l)	1.52 ± 0.39	2.06 ± 0.65	2.35 ± 0.43	< 0.0001	< 0.0001	0.04
ApoA-I (g/l)	1.63 ± 0.31	1.87 ± 0.42	1.98 ± 0.27	0.007	< 0.0001	0.11
LpAI (g/l)	0.51 ± 0.13	0.57 ± 0.20	0.75 ± 0.18	0.23	< 0.0001	0.001
LpAI:AII (g/l)	1.11 ± 0.20	1.30 ± 0.25	1.24 ± 0.18	0.003	0.02	0.18
ApoE (g/dl)	3.76 ± 0.97	4.12 ± 1.47	4.31 ± 1.88	0.36	0.16	0.67
Triglycerides (mmol/l)	1.23 ± 0.65	1.37 ± 0.98	0.83 ± 0.39	0.20	0.01	< 0.0001
LDL-C $(mmol/l)$	3.03 ± 1.07	2.80 ± 1.02	3.58 ± 0.99	0.27	0.13	0.03
ApoB (g/l)	1.16 ± 0.36	1.08 ± 0.40	1.12 ± 0.26	0.77	0.26	0.37
Total cholesterol (mmol/l)	5.07 ± 1.34	5.47 ± 1.18	6.31 ± 1.01	0.13	0.001	0.14

ApoA-I, apolipoprotein A-I; LpAI, high density lipoprotein containing apolipoprotein A-I only; LpAI:AII, high density lipoprotein containing apolipoprotein A-I and A-II. Values are given as means \pm SD. P1, level of significance, CETP-IVS7+1 heterozygotes versus family controls; P2, level of significance, HALP versus family controls; P3, level of significance, HALP versus CETP-IVS7+1 heterozygotes. Correction was performed for differences in age, sex, smoking, BMI, and statin use when these parameters contributed significantly to the model. Exclusion of the individual receiving fibrate therapy did not affect the outcome.

tween the groups, indicative of larger LDL particles in the HALP group. When comparing the CETP-IVS $7+1$ heterozygotes with controls, a slight reduction in LDL-C in the heterozygotes did not reach statistical significance, accompanied by the absence of differences in apoB concentration.

Thus, the CETP-IVS7+1 heterozygotes displayed an isolated high HDL-C phenotype that clearly differed from that of the HALP group, which exhibited a distinct high-HDL-C/low-triglyceride phenotype.

Other parameters

In the family with partial CETP deficiency, several additional analyses were carried out; these are summarized in Table 4. The CETP-IVS7 $+1$ mutation did not affect plasma LCAT activity (88 \pm 12 vs. 75 \pm 20%; P = 0.15) and phospholipid transfer protein activity (113 \pm 26 vs. 125 \pm 26% ; $P = 0.21$). Furthermore, NMR lipoprotein profiling in all 25 CETP-IVS7+1 heterozygotes and 20 family controls revealed increased HDL size in the heterozygotes $(9.6 \pm 0.6 \text{ vs. } 9.2 \pm 0.5 \text{ nm in controls}; P = 0.008)$. The observed increase of HDL-C levels could be mainly accounted for by the accumulation of cholesterol in the large HDL particles. The apparent absence of an effect of $CETP-IVS7+1$ on plasma triglyceride levels in heterozygotes compared with controls $(P = 0.20;$ Table 3) was reflected by similar triglyceride concentrations in large, intermediate, and small VLDLs $[25.0 \pm 27.6 \text{ vs. } 35.0 \pm 48.6 \text{ mg/d}]$ $(P = 0.20)$, 46.0 \pm 31.7 vs. 47.8 \pm 25.5 mg/dl ($P = 0.81$), and 8.5 ± 9.4 vs. 10.2 ± 6.8 mg/dl ($P = 0.70$), respectively]. A trend toward an increased LDL size was observed in the CETP IVS7 $+1$ heterozygotes compared with the controls $(P = 0.09)$; levels of small as well as large LDL particles were not affected ($P = 0.20$ and 0.55, respectively).

DISCUSSION

In contrast to studies on HALP in Japan, this study shows that plasma CETP activity and CETP concentration are not related to HDL-C levels in 95 unrelated Caucasians with HALP. Screening for CETP mutations in this cohort

led to the identification of a novel splice site defect (CETP- $IVS7+1$) in only one individual, indicating that CETP deficiency is rare among Caucasians of Dutch descent. Contrasting with the absence of a relation between CETP and HDL-C in the entire cohort, this CETP defect, causing a marked loss of CETP mass and activity, is associated with isolated high HDL-C in the family of this proband. This report describes for the first time in detail the effect of a CETPsplicesitemutationonlipidmetabolisminCaucasians.

CETP and HALP

Our data clearly indicate that high HDL-C levels in Caucasian men and women with HALP are unlikely to result from differences in plasma CETP activity or CETP concentration. In fact, we noted a complete absence of correlations between HDL-C and CETP activity, CETP concentration, or CETP specific activities in this study. These findings agree with those from other studies in Caucasians without HALP (31, 32). Furthermore, population-based studies determining the contributions of common CETP gene variants, known to be associated with decreased plasma CETP concentrations, demonstrated that these polymorphisms explain only a small proportion of the observed variations of HDL-C in Caucasians (1, 32). However, these findings contrast with reports that CETP explains a large portion of HALP in Japan (17). Together, the current data lend support to the notion that CETP is not a major determinant of increased HDL-C levels in the general Caucasian population, not even in the setting of extremely high levels of HDL-C.

CETP deficiency in Caucasians

This study indicates that genetic CETP deficiency is rare among subjects with HALP in The Netherlands. Only 1 of 95 HALP families was identified with partial genetic CETP deficiency. This novel defect is shown to cause skipping of exon 7 and is predicted to result in a premature truncation of the CETP protein, a product that could not be detected in plasma of the carriers using a dedicated ELISA.

The identification of this large family with partial CETP deficiency enabled us to analyze the effects of this disorder

TABLE 4. Lipoprotein-modifying enzymes and NMR lipoprotein profiling in heterozygotes for a novel CETP splice site mutation $(IVS7+1)$ and family controls

Variable	CETP-IVS7+1 Heterozygotes $(n = 25)$	Family Controls $(n = 25)$	\boldsymbol{P}
LCAT activity $(\%)$	88 ± 12	75 ± 20	0.15
Phospholipid transfer protein activity $(\%)$	113 ± 26	125 ± 26	0.21
HDL size (nm)	9.6 ± 0.6	9.2 ± 0.5	0.008
Large HDL $(\mu \text{mol}/l)$	13.7 ± 6.3	9.0 ± 4.9	0.003
Small HDL $(\mu \text{mol}/l)$	20.6 ± 3.9	22.8 ± 2.8	0.04
VLDL triglyceride content			
Large VLDL (mg/dl)	25.0 ± 27.6	35.0 ± 48.6	0.20
Intermediate VLDL (mg/dl)	46.0 ± 31.7	47.8 ± 25.5	0.81
Small VLDL (mg/dl)	8.5 ± 9.4	10.2 ± 6.8	0.70
LDL size (nm)	21.4 ± 0.9	21.0 ± 0.8	0.09
Large LDL $(\mu \text{mol}/l)$	472.2 ± 133.4	429.8 ± 228.7	0.55
Small LDL $(\mu \text{mol}/l)$	611.0 ± 443.7	810.6 ± 439.6	0.20

Values are given as means \pm SD; P values are for CETP-IVS7+1 heterozygotes versus family controls after correction for differences in age, sex, smoking, BMI, and statin use when these parameters contributed significantly to the model. None of the individuals received fibrate therapy.

on lipid metabolism in Caucasians and compare the effects with those in Japanese subjects with heterozygosity for another CETP splice site defect, Int $14+1$ G \rightarrow A (15). Compared with their family controls, heterozygotes for CETP-IVS7+1 or Int14+1 G \rightarrow A present with 50% and 40% reductions of CETP activity, respectively, and consequential increases of HDL-C (35% and 75%, respectively) (5). We have no explanation for the stronger effect on HDL-C in Japanese subjects, but this might relate to differences in lifestyle and genetic background. As seen in our $CETP-IVS7+1$ heterozygotes, the Japanese counterparts did not differ from their controls with respect to LDL-C, apoB, and triglyceride levels, further emphasizing an isolated high-HDL-C phenotype in partial CETP deficiency. Interestingly, these findings differ from those identified after inhibition of CETP activity using torcetrapib, a pharmaceutical CETP inhibitor (12). In that study, a less pronounced reduction of CETP activity $(-38%)$ significantly decreased baseline levels of LDL-C and triglycerides by -17% and -18% , respectively, in individuals with low HDL-C who had already received atorvastatin. In addition, the effects of torcetrapib on lipoprotein sizes were stronger compared with those observed in heterozygotes for CETP-IVS7+1 with a 50% loss of CETP activity. Also, less pronounced inhibition of CETP with JTT-705 in mildly dyslipidemic individuals (up to -37%) showed significant decrease of LDL-C levels (13). Apparently, the effects of lifelong endogenous loss of CETP as a result of CETP gene mutations cannot be compared directly with the loss of CETP activity induced by inhibitory compounds.

Partial CETP deficiency versus HALP without CETP deficiency

Comparing the heterozygotes for CETP-IVS $7+1$ with the remaining group of 94 individuals with HALP revealed some interesting differences. In contrast to the CETP-IVS711 heterozygotes, the HALP group exhibited a marked 40% lower mean triglyceride level, for which we have no direct explanation. Furthermore, HDL subfraction analyses by means of rocket gel electrophoresis indicated that the CETP-IVS7 $+1$ mutation favors the occurrence of LpAI:AII. There exists very little information in the current literature on this topic. In one other study of partial CETP deficiency, it was shown that LpAI and LpAI:AII levels were not different between heterozygous CETP-deficient subjects and controls (33), but only five affected individuals were studied. Regarding LpAI and LpAI:AII fractions and the risk of cardiovascular disease, data from epidemiological studies are inconsistent, reporting reductions of LpAI only (34, 35) as well as reductions of LpAI and LpAI:AII (36–38) in coronary heart disease patients. Very recently, Asztalos et al. (39) showed that neither LpAI nor LpAI:AII was significantly associated with coronary heart disease prevalence in The Framingham Offspring Study and the placebo group of the Veterans Affairs HDL Intervention Trial. Given the power of the latter studies ($n = 1,019$ and 741, respectively), it can be assumed that the different concentrations in the respective HDL subfractions in our study are unlikely to have an effect on atherosclerosis.

We also performed NMR lipoprotein subfraction analysis in the family members with partial CETP deficiency. Unfortunately, there exist no NMR data on HDL characteristics in genetic CETP deficiency in the literature. However, in agreement with studies using HPLC (18) or gel electrophoresis (19) techniques in CETP-deficient subjects, as well as observations from pharmacological CETP inhibition (12), we observed increases of mean HDL size and increased levels of large HDL particles. We also identified a trend ($P = 0.09$) toward increased LDL size in the CETP-IVS $7+1$ heterozygotes, as also observed in individuals treated with CETP inhibitors (12). Data on LDL size derived from genetically CETP-deficient Japanese subjects are limited to only a few studies with very small sample sizes $[n = 2 (20)$ and $n = 5 (40)$], again using techniques other than NMR. Given this paucity of data, as well as the observation of a trend toward statistical significance in our study, the effects of genetic CETP deficiency on LDLrelated NMR parameters need further investigation.

Regarding the clinical consequences of these findings, it is likely that the observed differences in lipid profiles of the CETP-IVS7 $+1$ heterozygotes may also translate into differences in the risk of cardiovascular disease. In this study, this is difficult to assess, given the small number of heterozygotes and their low average age $(39 \pm 22 \text{ years})$. Nevertheless, to investigate this, we measured carotid intima media thickness in 67 of 94 HALP individuals, in 19 heterozygotes for the CETP-IVS7 $+1$ mutation, and in 43 family controls (the latter group was expanded for this imaging study). The data show no statistically significant differences among the three groups (see supplementary Fig. II), but the limited number of individuals does not allow for firm conclusions.

Conclusions

This study shows that, overall, CETP does not play a major role in defining HALP in Caucasians of Dutch descent. However, heterozygosity for a novel CETP splice site mutation, identified in one family, was associated with an isolated high HDL-C level as the main lipid phenotype.

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REFERENCES

1. Boekholdt, S. M., O. W. Souverein, M. W. Tanck, G. K. Hovingh, J. A. Kuivenhoven, R. I. Peters, H. Jansen, P. M. Schiffers, E. E. van der Wall, P. A. Doevendans, et al. 2006. Common variants of multiple genes that control reverse cholesterol transport together ex-

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http://www.jlr.org/content/suppl/2007/01/17/M600405-JLR20
0.DC1.html

plain only a minor part of the variation of HDL cholesterol levels. Clin. Genet. 69: 263–270.

- 2. Rigotti, A., B. L. Trigatti, M. Penman, H. Rayburn, J. Herz, and M. Krieger. 1997. A targeted mutation in the murine gene encoding the high density lipoprotein (HDL) receptor scavenger receptor class B type I reveals its key role in HDL metabolism. Proc. Natl. Acad. Sci. USA. 94: 12610–12615.
- 3. Acton, S., D. Osgood, M. Donoghue, D. Corella, M. Pocovi, A. Cenarro, P. Mozas, J. Keilty, S. Squazzo, E. A. Woolf, et al. 1999. Association of polymorphisms at the SR-BI gene locus with plasma lipid levels and body mass index in a white population. Arterioscler. Thromb. Vasc. Biol. 19: 1734–1743.
- 4. Von Eckardstein, A., H. Holz, M. Sandkamp, W. Weng, H. Funke, and G. Assmann. 1991. Apolipoprotein C–III(Lys58-Glu). Identification of an apolipoprotein C–III variant in a family with hyperalphalipoproteinemia. J. Clin. Invest. 87: 1724–1731.
- 5. Nagano, M., S. Yamashita, K. Hirano, M. Takano, T. Maruyama, M. Ishihara, Y. Sagehashi, T. Kujiraoka, K. Tanaka, H. Hattori, et al. 2004. Molecular mechanisms of cholesteryl ester transfer protein deficiency in Japanese. J. Atheroscler. Thromb. 11: 110–121.
- 6. Drayna, D., A. S. Jarnagin, J. McLean, W. Henzel, W. Kohr, C. Fielding, and R. Lawn. 1987. Cloning and sequencing of human cholesteryl ester transfer protein cDNA. Nature. 327: 632–634.
- 7. Lusis, A. J., S. Zollman, R. S. Sparkes, I. Klisak, T. Mohandas, D. Drayna, and R. M. Lawn. 1987. Assignment of the human gene for cholesteryl ester transfer protein to chromosome 16q12–16q21. Genomics. 1: 232–235.
- 8. Agellon, L. B., E. M. Quinet, T. G. Gillette, D. T. Drayna, M. L. Brown, and A. R. Tall. 1990. Organization of the human cholesteryl ester transfer protein gene. Biochemistry. 29: 1372–1376.
- 9. Jarnagin, A. S., W. Kohr, and C. Fielding. 1987. Isolation and specificity of a Mr 74,000 cholesteryl ester transfer protein from human plasma. Proc. Natl. Acad. Sci. USA. 84: 1854–1857.
- 10. de Grooth, G. J., A. H. Klerkx, E. S. Stroes, A. F. Stalenhoef, J. J. Kastelein, and J. A. Kuivenhoven. 2004. A review of CETP and its relation to atherosclerosis. J. Lipid Res. 45: 1967–1974.
- 11. Clark, R. W., T. A. Sutfin, R. B. Ruggeri, A. T. Willauer, E. D. Sugarman, G. Magnus-Aryitey, P. G. Cosgrove, T. M. Sand, R. T. Wester, J. A. Williams, et al. 2004. Raising high-density lipoprotein in humans through inhibition of cholesteryl ester transfer protein: an initial multidose study of torcetrapib. Arterioscler. Thromb. Vasc. Biol. 24: 490–497.
- 12. Brousseau, M. E., E. J. Schaefer, M. L. Wolfe, L. T. Bloedon, A. G. Digenio, R. W. Clark, J. P. Mancuso, and D. J. Rader. 2004. Effects of an inhibitor of cholesteryl ester transfer protein on HDL cholesterol. N. Engl. J. Med. 350: 1505–1515.
- 13. de Grooth, G. J., J. A. Kuivenhoven, A. F. Stalenhoef, J. de Graaf, A. H. Zwinderman, J. L. Posma, A. van Tol, and J. J. Kastelein. 2002. Efficacy and safety of a novel cholesteryl ester transfer protein inhibitor, JTT-705, in humans: a randomized phase II doseresponse study. Circulation. 105: 2159–2165.
- 14. Kuivenhoven, J. A., G. J. de Grooth, H. Kawamura, A. H. Klerkx, F. Wilhelm, M. D. Trip, and J. J. Kastelein. 2005. Effectiveness of inhibition of cholesteryl ester transfer protein by JTT-705 in combination with pravastatin in type II dyslipidemia. Am. J. Cardiol. 95: 1085–1088.
- 15. Brown, M. L., A. Inazu, C. B. Hesler, L. B. Agellon, C. Mann, M. E. Whitlock, Y. L. Marcel, R. W. Milne, J. Koizumi, and H. Mabuchi. 1989. Molecular basis of lipid transfer protein deficiency in a family with increased high-density lipoproteins. Nature. 342: 448-451.
- 16. Takahashi, K., X. C. Jiang, N. Sakai, S. Yamashita, K. Hirano, H. Bujo, H. Yamazaki, J. Kusunoki, T. Miura, and P. Kussie. 1993. A missense mutation in the cholesteryl ester transfer protein gene with possible dominant effects on plasma high density lipoproteins. J. Clin. Invest. 92: 2060–2064.
- 17. Maruyama, T., N. Sakai, M. Ishigami, K. Hirano, T. Arai, S. Okada, E. Okuda, A. Ohya, N. Nakajima, K. Kadowaki, et al. 2003. Prevalence and phenotypic spectrum of cholesteryl ester transfer protein gene mutations in Japanese hyperalphalipoproteinemia. Atherosclerosis. 166: 177–185.
- 18. Yamashita, S., D. Y. Hui, J. R. Wetterau, D. L. Sprecher, J. A. Harmony, N. Sakai, Y. Matsuzawa, and S. Tarui. 1991. Characterization of plasma lipoproteins in patients heterozygous for human plasma cholesteryl ester transfer protein (CETP) deficiency: plasma CETP regulates high-density lipoprotein concentration and composition. Metabolism. 40: 756-763.
- 19. Arai, T., T. Tsukada, T. Murase, and K. Matsumoto. 2000. Particle

size analysis of high density lipoproteins in patients with genetic cholesteryl ester transfer protein deficiency. Clin. Chim. Acta. 301: 103–117.

- 20. Sakai, N., Y. Matsuzawa, K. Hirano, S. Yamashita, S. Nozaki, Y. Ueyama, M. Kubo, and S. Tarui. 1991. Detection of two species of low density lipoprotein particles in cholesteryl ester transfer protein deficiency. Arterioscler. Thromb. 11: 71–79.
- 21. Teh, E. M., P. J. Dolphin, W. C. Breckenridge, and M. H. Tan. 1998. Human plasma CETP deficiency: identification of a novel mutation in exon 9 of the CETP gene in a Caucasian subject from North America. J. Lipid Res. 39: 442–456.
- 22. Miettinen, H. E., H. Gylling, J. Tenhunen, J. Virtamo, M. Jauhiainen, J. K. Huttunen, I. Kantola, T. A. Miettinen, and K. Kontula. 1998. Molecular genetic study of Finns with hypoalphalipoproteinemia and hyperalphalipoproteinemia: a novel Gly230 Arg mutation (LCAT[Fin]) of lecithin:cholesterol acyltransferase (LCAT) accounts for 5% of cases with very low serum HDL cholesterol levels. Arterioscler. Thromb. Vasc. Biol. 18: 591–598.
- 23. Funke, H., H. Wiebusch, L. Fuer, S. Muntoni, H. Schulte, and G. Assmann. 1994. Identification of mutations in the cholesteryl ester transfer protein in Europeans with elevated high density lipoprotein cholesterol (Abstract). Circulation. 90: I-241.
- 24. Rhyne, J., M. J. Ryan, C. White, T. Chimonas, and M. Miller. 2006. The two novel CETP mutations Gln87X and Gln165X in a compound heterozygous state are associated with marked hyperalphalipoproteinemia and absence of significant coronary artery disease. J. Mol. Med. 84: 647–650.
- 25. Niemeijer-Kanters, S. D., G. M. Dallinga-Thie, F. C. Ruijter-Heijstek, A. Algra, D. W. Erkelens, J. D. Banga, and H. Jansen. 2001. Effect of intensive lipid-lowering strategy on low-density lipoprotein particle size in patients with type 2 diabetes mellitus. Atherosclerosis. 156: 209–216.
- 26. Dullaart, R. P., W. J. Sluiter, L. D. Dikkeschei, K. Hoogenberg, and A. van Tol. 1994. Effect of adiposity on plasma lipid transfer protein activities: a possible link between insulin resistance and high density lipoprotein metabolism. Eur. J. Clin. Invest. 24: 188–194.
- 27. Otvos, J. D., E. J. Jeyarajah, and W. C. Cromwell. 2002. Measurement issues related to lipoprotein heterogeneity. Am. J. Cardiol. 90: 22i–29i.
- 28. Boekholdt, S., J. Kuivenhoven, G. Hovingh, J. W. Jukema, J. Kastelein, and A. van Tol. 2004. CETP gene variation: relation to lipid parameters and cardiovascular risk. Curr. Opin. Lipidol. 15: 393–398.
- 29. Frisdal, E., A. Klerkx, W. Le Goff, M. W. Tanck, J. P. Lagarde, J. W. Jukema, J. Kastelein, M. J. Chapman, and M. Guerin. 2005. Functional interaction between $-629C/A$, $-971G/A$ and $-1337C/T$ polymorphisms in the CETP gene as a major determinant of promoter activity and plasma CETP concentration in the REGRESS Study. Hum. Mol. Genet. 14: 2607-2618.
- 30. Ritsch, A., C. Ebenbichler, E. Naschberger, W. Schgoer, U. Stanzl, H. Dietrich, P. C. Heinrich, K. Saito, and J. R. Patsch. 2004. Phagedisplayed recombinant single-chain antibody fragments with high affinity for cholesteryl ester transfer protein (CETP): cDNA cloning, characterization and CETP quantification. Clin. Chem. Lab. Med. 42: 247–255.
- 31. Fumeron, F., D. Betoulle, G. Luc, I. Behague, S. Ricard, O. Poirier, R. Jemaa, A. Evans, D. Arveiler, and P. Marques-Vidal. 1995. Alcohol intake modulates the effect of a polymorphism of the cholesteryl ester transfer protein gene on plasma high density lipoprotein and the risk of myocardial infarction. *J. Clin. Invest.* 96: 1664-1671.
- 32. Klerkx, A. H. E. M., M. W. T. Tanck, J. J. P. Kastelein, H. O. F. Molhuizen, J. W. Jukema, A. H. Zwinderman, and J. A. Kuivenhoven. 2003. Haplotype analysis of the CETP gene: not TaqIB, but the closely linked $-629C \rightarrow A$ polymorphism and a novel promoter variant are independently associated with CETP concentration. Hum. Mol. Genet. 12: 111–123.
- 33. Asztalos, B. F., K. V. Horvath, K. Kajinami, C. Nartsupha, C. E. Cox, M. Batista, E. J. Schaefer, A. Inazu, and H. Mabuchi. 2004. Apolipoprotein composition of HDL in cholesteryl ester transfer protein deficiency. J. Lipid Res. 45: 448–455.
- 34. Puchois, P., A. Kandoussi, P. Fievet, J. L. Fourrier, M. Bertrand, E. Koren, and J. C. Fruchart. 1987. Apolipoprotein A-I containing lipoproteins in coronary artery disease. Atherosclerosis. 68: 35–40.
- 35. Stampfer, M. J., F. M. Sacks, S. Salvini, W. C. Willett, and C. H. Hennekens. 1991. A prospective study of cholesterol, apolipoproteins, and the risk of myocardial infarction. N. Engl. J. Med. 325: 373–381.

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0.DC1.html

- 36. Genest, J. J., Jr., J. M. Bard, J. C. Fruchart, J. M. Ordovas, P. F. Wilson, and E. J. Schaefer. 1991. Plasma apolipoprotein A-I, A-II, B, E and C-III containing particles in men with premature coronary artery disease. Atherosclerosis. 90: 149–157.
- 37. Parra, H. J., D. Arveiler, A. E. Evans, J. P. Cambou, P. Amouyel, A. Bingham, D. McMaster, P. Schaffer, P. Douste-Blazy, and G. Luc. 1992. A case-control study of lipoprotein particles in two populations at contrasting risk for coronary heart disease. The ECTIM Study. Arterioscler. Thromb. 12: 701–707.
- 38. Buring, J. E., G. T. O'Connor, S. Z. Goldhaber, B. Rosner, P. N. Herbert, C. B. Blum, J. L. Breslow, and C. H. Hennekens. 1992. Decreased HDL2 and HDL3 cholesterol, Apo A-I and

Apo A-II, and increased risk of myocardial infarction. Circulation. 85: 22–29.

- 39. Asztalos, B. F., S. Demissie, L. A. Cupples, D. Collins, C. E. Cox, K. V. Horvath, H. E. Bloomfield, S. J. Robins, and E. J. Schaefer. 2006. LpA-I, LpA-I:A-II HDL and CHD-risk: the Framingham Offspring Study and the Veterans Affairs HDL Intervention Trial. Atherosclerosis. 188: 59–67.
- 40. Bisgaier, C. L., M. V. Siebenkas, M. L. Brown, A. Inazu, J. Koizumi, H. Mabuchi, and A. R. Tall. 1991. Familial cholesteryl ester transfer protein deficiency is associated with triglyceride-rich low density lipoproteins containing cholesteryl esters of probable intracellular origin. J. Lipid Res. 32: 21–33.

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